Total Synthesis of Balanol

K. C. Nicolaou,* Mark E. Bunnage, and Kazunori Koide Department of Chemistry, The Scripps Research Institute 10666 North Torrey Pines Road, La Jolla, California 92037 Department of Chemistry, University of California, San Diego 9500 Gilman Drive, La Jolla, California 92093 Received June 8, 1994

The recent isolation and structural elucidation of balanol (1, Figure 1),¹ an unusual metabolite produced by the fungus Verticillium balanoides, represents a significant new development in the quest for effective inhibitors of protein kinase C (PKC).² PKC-mediated signal transduction is known to lead to a variety of cellular responses, including gene expression and cell proliferation,^{2,3} and activated PKC has been implicated in conditions as diverse as cancer, cardiovascular disorders, asthma, inflammation, diabetes, CNS dysfunction, and HIV infection.³ Consequently, the identification of potent and selective PKC inhibitors not only may serve to further illuminate signal transduction pathways but also may result in the development of novel drugs with considerable therapeutic value.³ As part of a program directed toward the identification of effective inhibitors of PKC, we have completed, and present herein, the total synthesis of balanol.

A retrosynthetic analysis of balanol (Figure 1) reveals two distinct structural domains coupled through a central ester linkage. The hexahydroazepine-containing fragment can be further simplified by dissection of the amide moiety to reveal the p-hydroxybenzoic acid moiety, and the synthetic challenge posed by balanol thus reduces to an asymmetric synthesis of the central hexahydroazepine ring and the generation of the fully-functionalized benzophenone fragment.

Our synthetic approach to the hexahydroazepine fragment is presented in Scheme 1.4 Treatment of the readily available homochiral amino acid D-serine (2) with di-tert-butyl dicarbonate (Boc₂O) afforded the N-Boc protected amino acid, which was subsequently converted to the corresponding methyl ester and finally silvlated to afford the fully protected derivative 3 in quantitative overall yield. Reduction of 3 with excess DIBALH afforded the corresponding amino aldehyde, which was then treated with Brown's diisopinocampheylborane reagent,⁵ allyl- $B(^{1}Ipc)_{2}$, to afford the desired syn-amino alcohol 4 as a 12:1 mixture of diastereoisomers. The syn configuration of 4 was confirmed by desilylation using TBAF and conversion to the known acetonide 5.6

With the correct stereochemical disposition of the amino and hydroxyl stereogenic centers secured, the stage was now set to develop the hexahydroazepine ring system. Firstly, amino alcohol 4 was protected as the corresponding acetonide (68% yield from 3), which was then subjected to hydroboration using 9-BBN to furnish the alcohol 6 upon alkaline hydroperoxide workup (97% yield). Introduction of an azide moiety as an amino group surrogate was now achieved by initial mesylation of the hydroxyl

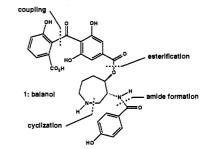
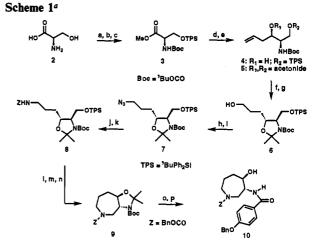


Figure 1.



^a Reagents and conditions: (a) 1.2 equiv of (Boc)₂O, 2.1 equiv of NaOH, 1,4-dioxane, H₂O, $0 \rightarrow 25 \,^{\circ}$ C, 2 h, 100%; (b) 1.1 equiv of K₂CO₃, 2.0 equiv of MeI, DMF, $0 \rightarrow 25$ °C, 100%; (c) 1.2 equiv of TPSC1, 1.4 equiv of imidazole, DMF, 25 °C, 14 h, 100%; (d) 2.5 equiv of DIBALH, toluene, -78 °C, 1.5 h; (e) 1.8 equiv of allyl-B(/Ipc)₂, Et₂O, -78 °C, 3.5 h; ethanolamine; (f) 5.0 equiv of 2,2-dimethoxypropane, CSA (1 mol %), CH₂Cl₂, 25 °C, 3 h, 68% (three steps); (g) 2.2 equiv of 9-BBN, THF, 20 h; NaOH, H₂O₂, 12 h, 97%; (h) 1.2 equiv of MsCl, 1.5 equiv of Et₃N, CH₂Cl₂, 0 °C, 10 min; (i) 8.0 equiv of NaN₃, DMF, 25 °C, 24 h, 98% (two steps); (j) H₂, Pd/C, THF, 19 h; (k) 1.1 equiv of benzyl chlorocarbonate, 3.0 equiv of NaOH, 1,4-dioxane, H2O, 0 °C, 15 min, 100% (two steps); (l) 1.2 equiv of TBAF, THF, 25 °C, 16 h, 96%; (m) 1.2 equiv of MsCl, 1.5 equiv of Et₃N, CH₂Cl₂, 0 °C, 20 min; (n) 1.2 equiv of KO^tBu added over 2 h, moderate dilution (0.02 M), THF, 25 °C, 80% (two steps); (o) excess TFA, CH₂Cl₂, 25 °C, 1 h; (p) 1.5 equiv of *p*-(benzyloxy)benzoyl chloride, 5.0 equiv of Et₃N, CH₂Cl₂, $0 \rightarrow 25 \text{ °C}$, 1.5 h, 73% (two steps).

moiety within 6 and subsequent sodium azide displacement to furnish 7 in 98% overall yield. Mild catalytic hydrogenolysis of the azide group over Pd/C and protection of the resultant amino moiety as its benzyloxycarbonyl (Z) derivative then provided 8 in quantitative yield overall. In order to facilitate closure to the hexahydroazepine ring, the primary alcohol within 8 was revealed by desilylation using TBAF (96% yield) and then activated toward displacement by formation of the corresponding mesylate. The 7-Exo-Tet cyclization was then effected by treatment with a slight excess of potassium tert-butoxide in THF at ambient temperature under moderately dilute conditions to afford the fully-protected hexahydroazepine fragment 9 in 80% yield. Finally, concomitant removal of the Boc and acetonide protecting groups within 9 by treatment with TFA, followed by in situ derivatization of the resultant free amino alcohol with p-(benzyloxy)benzoyl chloride, afforded the targeted fragment 10 in 73% overall yield (36% overall from 2).

With the hexahydroazepine fragment of balanol successfully secured, we anticipated that the remaining benzophenone domain could be prepared by utilization of a Stille-type coupling⁷ of the appropriately functionalized acid chloride and arylstannane

^{(1) (}a) Kulanthaivel, P.; Hallock, Y. F.; Boros, C.; Hamilton, S. M.; Janzen, W. P.; Ballas, L. M.; Loomis, C. R.; Jiang, J. B.; Katz, B.; Steiner, J. R.; Clardy, J. J. Am. Chem. Soc. 1993, 115, 6452. (b) This molecule has also been isolated from an alternative genus of fungi and termed "azepinostatin", see: Ohshima, S.; Yanagisawa, M.; Katoh, A.; Fujii, T.; Sano, T.; Matsukuma, S.; Furumai, T.; Fujiu, M.; Watanabe, K.; Yokose, K.; Arisawa, M.; Okuda, T. J. Antibiot. 1994, 47, 639.
 (2) (a) Newton, A. C Annu. Rev. Biophys. Biomol. Struct. 1993, 22, 1.

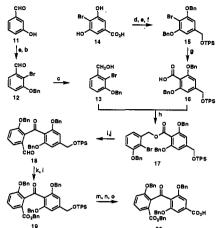
 ⁽b) Farago, A.; Nishizuka, Y. FEBS Lett. 1990, 268, 350. (c) Stabel, S.;
 Parker, P. J. Pharmacol. Ther. 1991, 51, 71. (d) Nishizuka, Y. Nature 1988, 334, 661. (e) Nishizuka, Y. Science 1986, 233, 305.
 (3) Bradshaw, D.; Hill, C. H.; Nixon, J. S.; Wilkinson, S. E. Agents Actions

^{1993. 38. 135.}

⁽⁴⁾ For a related approach to this fragment, see: Müller, A.; Takyar, D. K.; Witt, S.; König, W. A. Liebigs Ann. Chem. 1993, 651.

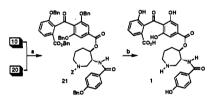
⁽⁵⁾ Brown, H. C.; Bhat, K. S.; Randad, R. S. J. Org. Chem. 1989, 54, 1570. (6) Hafner, A.; Duthaler, R. O.; Marti, R.; Rihs, G.; Rothe-Streit, P.; Schwarzenbach, F. J. Am. Chem. Soc. 1992, 114, 2321.

Scheme 2^a



^a Reagents and conditions: (a) 1.0 equiv of BnBr, 1.5 equiv of K₂CO₃, DMF, 25 °C, 12 h, 100%; (b) 1.03 equiv of "BuLi, 1.07 equiv of MeNHCH₂CH₂NMe₂, PhH, $0 \rightarrow 25$ °C, 0.5 h; aldehyde, $0 \rightarrow 25$ °C, 0.5 h; 3.0 equiv of PhLi, $0 \rightarrow 25$ °C, 7.5 h; 4.0 equiv of CF₃BrCF₃Br, THF, -78 → 25 °C, 12 h; 10% aqueous HCl, 0 °C, 77%; (c) 1.2 equiv of DIBALH, CH₂Cl₂, $-78 \rightarrow 25$ °C, 1 h, 93%; (d) 3.3 equiv of BnBr, 5.0 equiv of K₂CO₃, DMF, 25 °C, 4 h, 96%; (e) 2.4 equiv of DIBALH, CH_2Cl_2 , -78 \rightarrow 0 °C, 1 h, 100%; (f) 1.2 equiv of TPSCl, 1.5 equiv of imidazole, DMF, 25 °C, 1 h, 82%; (g) 1.3 equiv of "BuLi, THF, -78 °C, 0.5 h; excess CO₂, $-78 \rightarrow 25$ °C, 0.5 h; 10% aqueous HCl, 60%; (h) 1.1 equiv of DEAD, 1.1 equiv of Ph₃P, THF, $0 \rightarrow 25$ °C, 40 min; 93%; (i) 1.0 equiv of "BuLi, THF, $-97 \rightarrow -78$ °C, 50 min; NH₄Cl (aqueous); (j) 1.5 equiv of NMO, cat TPAP, CH₃CN, 25 °C, 0.5 h, 74% (two steps); (k) 3.0 equiv of NaClO₂, 3.0 equiv of NaH₂PO₄, excess 2-methyl-2butene, THF, 'BuOH, H₂O, 25 °C, 8 h; (1) 2.0 equiv of BnBr, 3.0 equiv of K₂CO₃, DMF, 25 °C, 1 h, 98% (two steps); (m) 1.5 equiv of TBAF, THF, 25 °C, 10 min, 95%; (n) 1.5 equiv of NMO, cat TPAP, CH₃CN, 25 °C, 0.5 h, 70%; (o) as in k, 1 h, 95%.

Scheme 3^a



^a Reagents and conditions: (a) 1.0 equiv of 10, 1.0 equiv of 20, 1.3 equiv of 2-chloro-1-methylpyridinium iodide, 0.5 equiv of DMAP, 20 equiv of Et₃N, CH₂Cl₂, 25 °C, 5 h, 79%; (b) 1 atm of H₂, Pd black, THF/H₂O/AcOH (16:4:1), 25 °C, 8 h, 100%.

components suggested by the disconnection in Figure 1. Unfortunately, such an approach was not rewarded with success and alternative procedures for the generation of this fragment were consequently sought. In particular, model studies⁸ demonstrated that lithium-halogen exchange of esters analogous to 17 (vide infra) was followed by intramolecular attack of the corresponding aryllithium species at the ester moiety.⁹ thereby generating a ketone linkage between the two aromatic components. The successful utilization of this approach in the synthesis of the benzophenone domain of balanol is outlined in Scheme 2.

The alcohol component of ester 17 was prepared from m-hydroxybenzaldehyde (11) by a three-step sequence commencing with the benzylation of the hydroxyl moiety using benzyl bromide/potassium carbonate in DMF (100% yield). Regiospecific lithiation of the resultant aldehyde, 10 followed by electrophilic bromination of the corresponding aryllithium,¹¹ then afforded 12 as a single regioisomer in 77% yield, and DIBALH reduction of this aldehyde furnished the desired benzyl alcohol 13 (93% yield). Commercially-available acid 14 was selected as a convenient precursor for the acid component of ester 17. Thus, tribenzylation of 14 using benzyl bromide/potassium carbonate in DMF occurred smoothly (96% yield), and the resultant benzyl ester was reduced using an excess of DIBALH to the corresponding primary alcohol (100% yield), which was then silvlated to afford 15 in 81% yield. With 15 in hand, we successfully secured the desired acid 16 in good yield (60%) via lithium-halogen exchange and trapping of the resulting aryllithium with carbon dioxide.

The coupling of 16 and 13 was most efficiently achieved using the Mitsunobu protocol,¹² which furnished the target ester 17 in 93% yield. The aryllithium species resulting from the low temperature (-100 °C) treatment of 17 with butyllithium successfully underwent the desired rearrangement to afford, upon protic workup, the corresponding alcohol, which could be isolated but was more conveniently oxidized directly to the aldehyde 18 (74% yield from 17) using TPAP-NMO.¹³ With this material in hand, sodium chlorite oxidation¹⁴ afforded the carboxylic acid, which was readily converted to the benzyl ester 19 in the normal manner (98% yield from 18). Subsequent TBAF-mediated desilylation of 19 revealed the corresponding alcohol (95% yield), which was in turn subjected to TPAP-NMO oxidation to afford the aldehyde (70% yield). Finally, oxidation of this aldehyde using sodium chlorite proceeded in excellent yield to furnish the fully-benzylated balanol benzophenone component 20.

The union of the two protected components 10 and 20 was successfully accomplished via esterification using the Mukaiyama procedure¹⁵ to afford the fully-protected balanol progenitor 21 in 79% yield (Scheme 3). The adoption of benzyl-derived protecting groups for the latent funtionalities in this coupling procedure was now rewarded since mild palladium-catalyzed hydrogenolysis of 21 resulted in the liberation of balanol (1) as a yellow solid in quantitative yield (>90% purity). Final purification by reverse-phase HPLC afforded a sample of balanol which exhibited characterization data consistent with the proposed structure and a ¹H NMR spectrum which correlated precisely with that of an authentic sample.¹⁶

It is noteworthy that the aforementioned synthetic strategies for the hexahydroazepine and the benzophenone fragments are particularly suitable for analog generation, and in order to elucidate the structural requirements for PKC inhibition, both the synthesis and biological evaluation of a variety of new balanoids are currently under investigation.

Acknowledgment. We would like to thank Dr. Palaniappan Kulanthaivel of Sphinx Pharmaceuticals for kindly providing copies of their NMR spectra for balanol and for useful discussions. M.E.B. would like to thank the EPSRC, U.K. (formerly SERC), for a NATO Postdoctoral Fellowship. This work was also financially supported by the National Institutes of Health and the Scripps Research Institute.

Supplementary Material Available: Characterization data for compounds 10, 20, 21, and 1 (5 pages). This material is contained in many libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.

^{(7) (}a) Labadie, J. W.; Tueting, D.; Stille, J. K. J. Org. Chem. 1983, 48, 4634. (b) Milstein, D.; Stille, J. K. J. Org. Chem. 1979, 44, 1613.

⁽⁸⁾ These studies will be described in the full account of this work

⁽⁹⁾ For an analogous rearrangement, see: Horne, S.; Rodrigo, R. J. Chem. Soc., Chem. Commun. 1992, 164.

¹⁰⁾ Comins, D. L.; Brown, J. D. J. Org. Chem. 1989, 54, 3730. (11) Moorlag, H.; Meyers, A. I. Tetrahedron Lett. 1993, 34, 6993.

⁽¹²⁾ Mitsunobu, O. Synthesis 1981, 1.
(13) Griffith, W. P.; Ley, S. V. Aldrichimica Acta 1990, 23, 13.
(14) Bal, B. S.; Childers, W. E.; Pinnick, H. W. Tetrahedron 1981, 37,

^{2091.}

⁽¹⁵⁾ Mukaiyama, T. Angew. Chem., Int. Ed. Engl. 1979, 18, 707. (16) NMR spectra for authentic balanol were kindly provided by Sphinx Pharmaceuticals.